

Biochemistry and Cell Biology of Silica Formation in Sponges

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ABSTRACT The main inorganic material forming the skeletal elements in Demospongiae as well as in Hexactinellida, the spicules, is amorphous silica. The spicules occur in the cytoplasm and the extracellular space and also in the nucleus (as silicate crystals) of some sponge cells; the function in the latter compartment is unknown. Recent evidence shows that the formation of spicules is mediated by the enzyme silicatein. The cDNA as well as the gene encoding this enzyme was cloned from *Suberites domuncula*. The recombinant silicatein catalyzes the synthesis of amorphous silicate using tetraethoxysilane as substrate. The enzyme is dependent on ferric iron. Silicatein also has proteolytic (cathepsin-like) activity. Incubation of primmorphs, a special form of 3D-cell aggregates, with silicon resulted in a strong increase of their size from 1–7 mm to ~10 mm. The morphogenetic activity of silicate is underscored by the finding that this ion increases gene expression of silicatein and collagen. Based on these findings, it is concluded that both iron and silicate stimulate the activity of silicatein. Furthermore, it is proposed that the growing spicules are surrounded by the scavenger receptor which might be considered as a docking molecule for the collagen matrix into which the spicules are embedded.

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INTRODUCTION

Sponges (Porifera) represent the oldest extant metazoan phylum (Müller, 1995, 2001). The earliest fossil records indicate that soft-bodied, presumably multicellular animals, including sponges (from the Ediacara fauna), appeared ~600 Myr ago (MYA) (Mehl et al., 1998; Brasier, 2000). However, the paleontological evidence for possible metazoan fossils is still under debate, like the recent claims about metazoan embryos from the Doushanto Formation (Xiao et al., 1999), which had been attributed to *Volvox*-like chlorophytes (Xue et al., 1998). However, molecular evidence appears to be reliable; based on analyses of informative protein sequences it had been proposed that the phylum Porifera evolved more than 600 MYA from the hypothetical Urmetazoa (reviewed in Müller, 1998). Furthermore, molecular biology techniques also provide evidence that, among the three classes within the phylum Porifera, the Hexactinellida evolved first, followed by the Demospongiae, and finally the Calcarea (reviewed in Müller et al., 1998). In this context it is notable that the Hexactinellida and the Demospongiae contain siliceous spicules as skeletal elements, while the spicules of the Calcarea are of calcareous material. Isotopic data revealed that siliceous biomineralization preceded calcareous biomineralization by more than 10 Myr (Brasier et al., 1997), thus further supporting this assumption.

It is surprising that during the Cambrian/Precambrian transition an explosive evolution of the biota occurred, characterized by the appearance of organisms with phosphate and also siliceous skeletons. During the Cambrian/Precambrian period, a stepwise increase in oxygen occurred (Brasier, 2000); this process

was followed by a depletion of dissolved iron in the marine environment. In the present day oceans, the seawater contains 1 nM of dissolved iron and <3 to 150 µM of dissolved silicon (silicate) (Kennish, 1994). The surface water is even poorer in silicon, with concentrations often below 3 µM (Nelson et al., 1995). In contrast, the concentration of silicon in human blood is ~140 µM and that of iron ~15 µM (Geigy Scientific Tables, 1984).

Recent studies reported that the expression of the gene for silicatein, which has been assumed to be the key enzyme for the synthesis of spicules in demosponges *Tethya aurantium* (Shimizu et al., 1998 as *T. aurantia*) and *Suberites domuncula* (Krasko et al., 2000), is controlled by extracellular silicate (Krasko et al., 2000). At ~1 µM of soluble silicate in seawater, the expression of silicatein is low; at 60 µM of silicate expression increases dramatically (Krasko et al., 2000). Moreover, extracellular silicate also acts on a second gene which is involved in spicule formation, the collagen gene (Krasko et al., 2000). The expression of this latter is further upregulated when sponge cells are incubated with an extracellular signaling factor, the

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cardiac myotrophin-related molecule, isolated from *S. domuncula* (Schröder et al., 2000).

We established the growth conditions for cells from sponges *in vitro* (Custodio et al., 1998; Müller et al., 1999). The 3D-aggregates formed were termed primmorphs. After dissociation, the cells start to proliferate (proliferation phase) when they are brought into contact with adjacent cells (Custodio et al., 1998; Müller et al., 1999). During this process, the spicules which contribute to control of morphogenesis are formed (Schröder et al., 2000; Krasko et al., 2000).

SILICA FORMATION: SILICATEIN Silicatein Sequence From *S. domuncula*

Protein. The *S. domuncula* cDNA (termed: *SUBDO-SILICA*) encoding the sponge silicatein was isolated and characterized (Krasko et al., 2000). The 1,169 bp sequence has an open reading frame of 993 nt; the predicted translation product of 330 amino acids (aa), *SILICA_SUBDO*, (Fig. 1A) has a calculated M_r of 36,306 Da.

As outlined before (Shimizu et al., 1998; Krasko et al., 2000), silicatein is a new member of the cathepsin subfamily. The *S. domuncula* silicatein shares 70% identical and 79% similar (identical plus physicochemical-related aa) aa with the *T. aurantium/aurantia* silicatein polypeptide (Shimizu et al., 1998); while an alignment with the corresponding *Geodia cydonium* cathepsin protein (Krasko et al., 1997) shows a lower identity/similarity of 43%/59%. The three amino acids Cys, His, and Asn, which form the catalytic triad of cysteine proteases, are present in the sponge cathepsin at the characteristic sites: Cys₁₂₅, His₁₆₄, and Asn₁₈₄ (Krasko et al., 2000). As reported before for the *T. aurantium* silicatein (Shimizu et al., 1998), the cluster comprising the characteristic hydroxy amino acids (serine) is also present in the corresponding *S. domuncula* molecule (Fig. 1A).

A processing step is required for the formation of active cathepsins, which is mediated either by autolysis or by a second protease. Based on published comparisons with cathepsins, the characteristic cleavage site is also present in the sponge silicatein sequences, suggesting a processing of a proenzyme with an M_r of 36,306 to the mature form with an M_r of 23,125 (Fig. 1A).

Phylogenetic Analysis. The highest similarity of the sponge silicatein is shared with the sponge cathepsin of the cathepsin-L type. This conclusion is drawn from the alignment (Fig. 1A) and the resulting unrooted tree (Fig. 1B). The *S. domuncula* silicatein branches off from the same root which also comprises the cathepsin-L from human (Fig. 1B). The other known human cathepsin molecules are more distantly related to the silicatein/cathepsin-L molecules; the sponges also contain other cathepsins, e.g., cathepsin B, as indicated in Figure 1.

Silicatein Gene. The complete gene from which the cDNA for silicatein derives was cloned from the cDNA library of *S. domuncula*. The total length of the gene is 2,280 nt (Fig. 2). It comprises six introns which are—as is usual for sponge sequences—short (Gamulin et al., 1997); the intron borders follow the “GT–AG” rule. Intron number 2 has 166 nt and intron number 5 (the longest) comprises 375 nt (Fig. 2). The cathepsin-spe-

cific motifs 1 to -3 are not interrupted by introns. Motif 1 is present between intron-2 and -3, motif 2 between and intron-5 and -6 and motif 3 is also in this gene segment.

Recombinant Silicatein. As recently reported (Krasko et al., 2002), the sequence including the putative catalytic segment, from aa₁₇ to aa₃₃₀ (nt₄₉ to nt₉₉₃), was used for expression in *Escherichia coli*. The cDNA was inserted into the bacterial oligohistidine expression vector pQE-30 (Qiagen, Chatsworth, CA) as summarized in Figure 3A. *E. coli* strain XL1-blue was transformed with this plasmid and expression of fusion protein was induced for 6 hours at 37°C with 1 mM isopropyl 1-thio-β-D-galactopyranoside (IPTG). The fusion protein was purified. The purity of the material was checked by electrophoresis on 10% polyacrylamide gels containing 0.1% sodium dodecyl sulfate. The purified silicatein, termed r-silicatein, was digested with 5 U of recombinant enterokinase. The recombinant silicatein was cleaved from the His-tag by digestion with recombinant enterokinase; the purified long form of the r-silicatein preparations had an M_r of 35 kDa (calculated size: 34,663) and the short form of M_r 26 kDa (22,149) (Fig. 3B, lane a). These two recombinant r-silicatein preparations were used for analysis of enzyme activity.

Silicatein Enzyme Assay. In a total volume of 500 μl, 100 μg of r-silicatein was added to the 25 mM Tris-HCl buffer (pH 7.2; 10 mM of DL-dithiothreitol, 100 mM NaCl). As substrate, 4.5 mM tetraethoxysilane was used. Where indicated, 5 mM MgCl₂ and/or 5 mM (final concentration) of Fe(++) citrate was added. The reaction was performed at 22°C for 90 minutes and terminated by centrifugation (20,000g; 4°C, 15 min). The sediment obtained was washed three times in ethanol and an aliquot of 250 μl was determined for released silicic acid using the “Silicon Test” colorimetric assay kit (Merck, Darmstadt, Germany; 1.14794). Based on the absorbance values at 810 nm, the absolute amounts of silicic acid were calculated after construction of a calibration curve using a silicon standard (Merck; 1.09947).

Enzymatic Activity: Silicate Formation. Both forms of the recombinant silicatein (r-silicatein) were found to increase the amount of polymerized silica significantly from 2.2 (assays without enzyme) to 5.9 ng/mg for the long form of the enzyme (2.4–6.1 ng/mg for the short form) compared to the assays without r-silicatein. If ferric iron [Fe(++)] was added to the reaction mixture, a 3-fold increase of the reaction product was observed in both the assay with the long form of the enzyme and that with the short form. In contrast, Mg(++) had no effect on the reaction (Fig. 4).

These experiments and those already published (Krasko et al., 2000, 2002; Müller et al., 2002), together with those reported for the *T. aurantium* model (Shimizu et al., 1998), provide strong evidence that silicatein is the major enzyme involved in silicate production required for the formation of spicules.

Enzymatic Activity: Proteolytic Activity. As outlined above (Fig. 1), the deduced protein sequence of the sponge silicatein shows high sequence similarity to lysosomal enzymes, the cathepsins. We therefore explored whether the recombinant silicatein also has proteolytic activity, in addition to the ability to form amor-

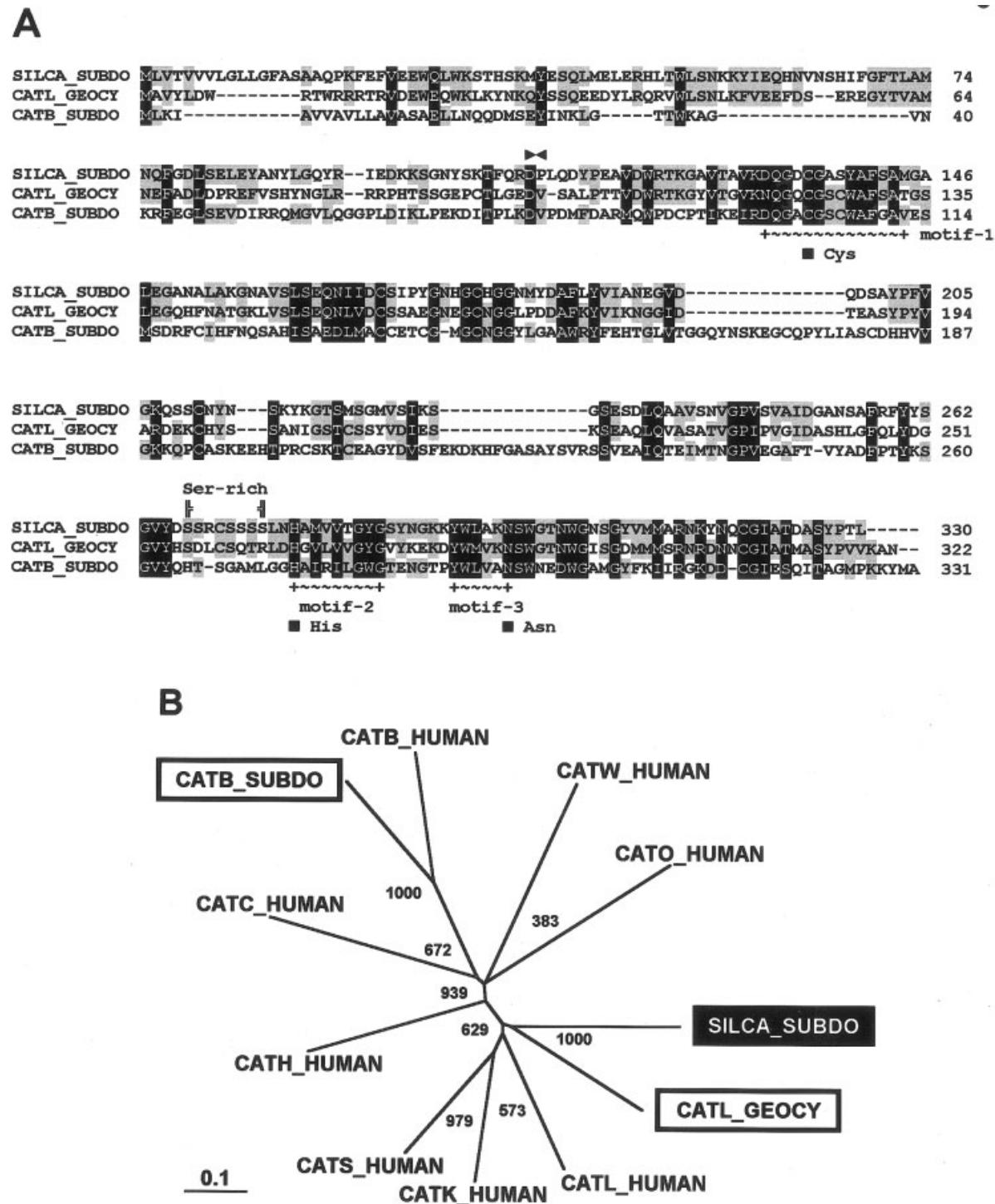


Fig. 1. Silicatein polypeptide. **A:** The deduced aa sequence from the *S. domuncula* cDNA encoding silicatein (termed SILICA_SUBDO) was aligned with the cathepsin sequences from sponges; the *S. domuncula* cathepsin B (CATB_SUBDO) and the *G. cydonium* cathepsin L (CATL_GEOCY, accession number Y10527). Residues conserved (similar or related with respect to their physicochemical properties) in all sequences are shown in white on black and those in at least two sequences in black on gray. The characteristic three active sites aa are indicated (■), the cleavage site for the processing to the active enzymes is indicated (►). Furthermore, the three motifs, motif-1 to -3, are highlighted. The serine-rich section in the silicatein sequence is

delimited above the alignment. **B:** These deduced sponge protein sequences were aligned with the different human cathepsin sequences: cathepsin B (CATB_HUMAN; P07858), cathepsin C (CATC_HUMAN; P53634), cathepsin H (CATH_HUMAN; P09668), cathepsin K (CATK_HUMAN; P43235), cathepsin L (CATL_HUMAN; P07711), cathepsin O (CATO_HUMAN; P43234), cathepsin S (CATS_HUMAN; P25774), and cathepsin W (CATW_HUMAN; P56202). The phylogenetic relationships of the sponge sequences within the human sequences is given in an unrooted tree. Scale bar indicates an evolutionary distance of 0.1 aa substitutions per position in the sequence.

CTAACTAAAT	TCTATACTGT	TTGTTATCAT	CCTTTATCAT	ATACATGCAC	TCACACAGGA	60
M L V	T V V	V L G L	L G F	A S A	A Q P K	20
CAATGCTTGT	CACAGTGGTA	GTACTGGTC	TACTGGGTT	TGCTTCTGCA	GCCCAGCCCA	120
F E F	V E E	W Q L W	K S T	H S K	M Y E S	40
AGTTTGAATT	TGTAGAAGAA	TGGCAGCTGT	GGAAGTCCAC	TCACTCTAAC	ATGTACGAGT	180
Q L M	E L E	R H L T	W L S	N K K	Y I E Q	60
CACAGTTAAT	GGAACTCGAA	AGACATCTGA	CGTGGCTCTC	CAATAAGAAA	TATATCGAGC	240
H N V	N S H	I F G F	T L A	M N Q	F G D L	80
AACACAATGT	CAACTCACAC	ATTTTCGGTT	TTACTCTGGC	AATGAACCAAG	TTGGAGATC	300
intron-1						
TGGTAAGATT	GGCATACTAT	TGTAGCATAA	TAATTATACA	AACTCCATGC	ATATATAAGT	360
ATAGGCTATA	TATAGGTCTA	ATTTAAGAGA	TTTGGTTGTG	ACAACATTTG	AAAAACTTGT	420
CATGAAGGCT	CAATGTGTAT	GAACCACCAA	TTCCAATCTT	ACGTATCCTT	TACTGTATAA	480
intron-1 S E L E Y A D Y L G 90						
TACCAAGCATT	CACCATATT	ATTGTTTTAG	AGTGAATTGG	AGTATGCTGA	CTATCTTGGC	540
Q Y R I	E D K	K S G	N Y S K	T F Q	R D P	110
CAGTATCGCA	TTGAGGATAA	AAAATCTGGC	AACTACTCAA	AGACTTTCA	GGGTGATCCT	600
L Q D Y	P E A	V D W	R T K G	A V T	A V K	130
CTACAGGACT	ACCCCTGAAGC	TGTAGACTGG	AGAACCAAAG	GAGCTGTCAAC	GGCTGTCAAG	660
D Q intron-2						132
GACCAGGTAT	TCTTCATAAA	AATGATTATT	CTTTCTATGC	ATGGAAACCT	GAATTGTTAA	720
TATCACCGAGC	CCAGTAAGCA	TATCTAAAGG	CAGCTTTCAT	TTGTACAACG	ATTTATAGGG	780
intron-2 G D C 135						
TGAATCTAGT	CATATTCAT	ATTTGAAACT	CATTAATGAC	GTATCAATCC	AGGGAGACTG	840
G A S	Y A F S	A M G	A L E	G A N A	L A K	155
TGGTGCTAGC	TATGCTTTA	GTGCTATGGG	TGCTTTGGAG	GGTGTCAATG	CTTTAGCCAA	900
G N A	V S L S	E Q N	I I D	C S I intron-3		171
GGGAAATGCA	GTATCTCTCA	GTGAACAGAA	CATCATTGAT	TGCTCGAGTT	AGTTCTATAA	960
GACGGTGACC	ACATGCATAC	GATAAGTTAT	TTCCATTGTT	ATAACTTTCG	AGGTCTATG	1020
TATTTATATA	CATAGATTG	GTGATGGAT	ATAATTATTG	TTTGTGTTTA	TCATTGACCA	1080
Int3 P	Y G N H	G C H	G G N	M Y D A	F L Y	189
CGCAGTTCT	TACGGTAACC	ACGGTTGTCA	TGGAGGCAAT	ATGTATGATG	CTTTTTGTA	1140
V I A	N E G V	D Q D	S A Y	P F V G	K in-4	207
TGTCATCGCT	AACGAGGGGG	TCGATCAGGA	CAGTGCATAT	CCATTGTTG	GAAAGGTGAA	1200
TAATAATTAT	TCTTCATCAC	AACAGTAATT	ATCGAACAGT	GTCCTAATAA	ACCAACTGTA	1260
ATGTCTGCAA	CTCTTAGAAT	TACCTATTCT	GTTGTTGTG	TTGTTGTTT	TTTGTTGTTG	1320
intron-4 Q S S C 211						
TTGTGTTGT	AGACTATGTT	TACTTTAGCC	ACTTCCCTGC	ACACACAGCA	ATCCAGCTGC	1380
N Y N S	K Y K	G T S	M S G M	V S I	K S G	231
AACTATAACAA	GTAAATACAA	AGGTACATCA	ATGTCGGGTA	TGGTGTCAAT	CAAAAGTGGT	1440
S E S D	L Q A	A V S	N V G P	V S V	A I D	251
AGTGAGTCTG	ACTTACAAGC	AGCTGTTCA	AACGTTGGCC	CTGTATCTGT	TGCTATTGAT	1500
G A N S	A F R	intron-5				258
GGTGCTAACAA	GTGCCCTCAG	GGTAAGACAG	ACTCACTCTA	ATTTAGTAAT	TAGCTGTTAG	1560
CGTATGGTTG	CGCTTACTTA	ATGTTGCT	TAATTACTTT	CCACTCTACT	ATTAATGACG	1620
CCGACTTTTC	CACGCATGTC	AGTTACAAAA	GAAGACGTTT	TCACATTGTC	TTGGGCAAT	1680
TTGATAGTTA	TTGCCACCAAG	TCGCTATTCA	GGCATGAAA	GAAGCACAAA	AAAGCCTATA	1740
CTATTGTTTC	TAATTCATG	CTTTTCGAA	TGCATGTATG	CCATGAATTG	AACATGACTG	1800
TCTATTGCTT	TGTGTACATA	GTGTATCCGT	AGGTTTCATAC	GTAAATTGTC	TTTGATTGATG	1860
intron-5 F Y Y S G V Y D 266						
TGGAGGTCT	TGTTCTGTT	ATACACATAC	ATACAGTTT	ACTACAGTGG	TGTCTATGAC	1920
S S R C	S S S	S L N	H A M V	V T G	Y G S	286
TCATCACGAT	GCTCTAGTAG	TAGTCTTAAC	CACGCAATGG	TAGTCACTGG	ATACGGATCA	1980
Y N G K	K Y W	L A K	N S intron-6			298
TACAATGGGA	AAAAATACTG	GCTGGCCAAG	AATAGGTTCG	CTCAGCTGTC	ACATATTGTT	2040
CTTGGGTTGC	CCCCCTCTCAC	TAATTACACAC	ACATATACAG	CATATACAGC	ATCACTCACC	2100
CGCATGTACA	CACTCACATT	CAATATGCCG	ATTGAGACTT	TTACCTCTT	ATATCTGACT	2160
intron-6 W G T N W G N S G Y V M M A R N 314						
CATACACACA	GCTGGGAAAC	TAACTGGGTT	AACAGTGGCT	ATGTGATGAT	GGCCCGCAAC	2220
K Y N Q	C G I	A T D	A S Y P	T L -		330
AAGTACAACC	AGTGTGGAAT	TGCTACCGAT	GCATCTTATC	CCACCCATA	AACTTATATA	2280

Fig. 2. Gene organization of silicatein. The deduced translation of the nt sequence into the polypeptide is given above the nucleotides. The borders of the introns are indicated.

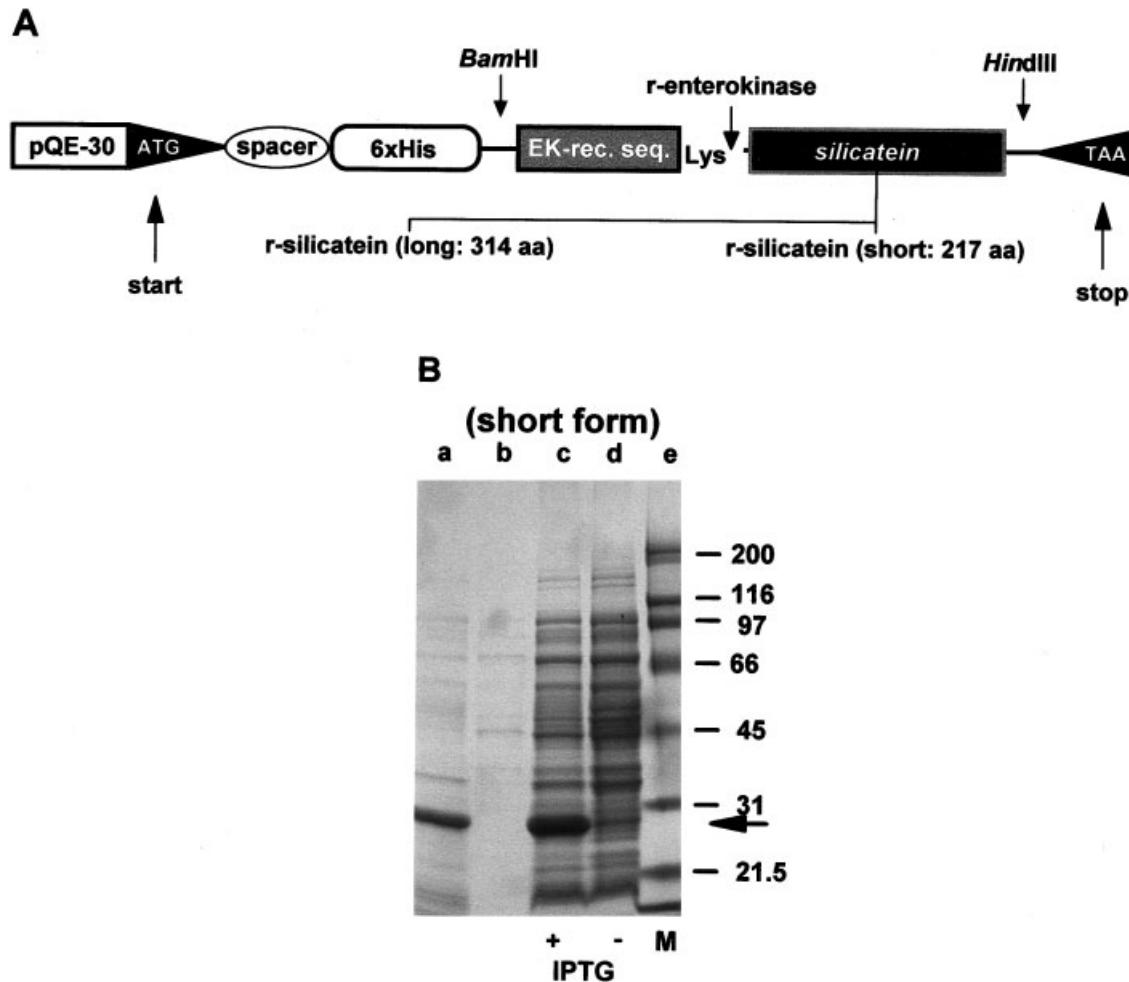


Fig. 3. Preparation of the recombinant silicatein. **A:** The *S. pombe* silicatein was inserted into the expression vector pQE-30 via the *Bam*HI (5'-end) and the *Hind*III (3'-end). Between oligohistidine (6xHis) and silicatein a nucleotide stretch corresponding the enterokinase recognition site [EK-rec. seq.] with the cleavage terminus [Lys ↓] was inserted. The start ATG as well as the stop TAA codon for transcription are indicated. The position at which the enterokinase cleaved the recombinant fusion protein is indicated. Two constructs of silicatein cDNA have been inserted: For the preparation of the long form of silicatein (comprising aa₁₇ to aa₃₃₀), the stretch spanning nt₄₉-nt₉₉₃ was selected, while for the short form of silicatein (aa₁₁₃ to

aa₃₃₀) the nt₃₃₇-nt₉₉₃ part of the sequence was used (Fig. 1A). **B:** Expression of the His-silicatein fusion protein. The expression of the short form of silicatein is shown. Aliquots of lysates from bacteria, which remained noninduced (-IPTG; lane d) or had been induced with IPTG (+IPTG; lane c) as well as the purified fusion protein (lane a) were analyzed by NaDODSO₄/PAGE (15% gels) as described in text. In lane b the fraction which had not been adsorbed to the Ni-NTA-agarose resin was analyzed. The fusion protein was subjected to digestion with recombinant enterokinase. The arrow points to the band corresponding to the recombinant silicatein protein. Marker proteins (M) were run in parallel.

phous silicate. To examine whether r-silicatein hydrolyses protein, the in situ activity after separation of the protein by NaDODSO₄/PAGE was monitored. Gelatin was used as substrate for the detection of enzymatic activity.

The short form of r-silicatein was separated by NaDODSO₄/PAGE using 10% polyacrylamide gels and 0.1% NaDODSO₄ according to Laemmli (1970). Bacterial extracts, obtained from *E. coli* which had been induced with IPTG (Fig. 3B, lane c) or had not been induced (Fig. 3B, lane d), were size-separated. Two types of gels were used: 1) a gel which did not contain gelatin (Fig. 5A) or 2) a gel which contained 0.1% gelatin (Fig. 5B). The gel without gelatin was immediately stained with Coomassie brilliant blue (Fig. 5A) to demonstrate the appearance of the recombinant sili-

catein in the extracts from IPTG-treated *E. coli* (Fig. 5A, lane a). In parallel, a zymogram was prepared under the same gel separation conditions using a gel which was supplemented with 0.1% gelatin (Fig. 5B); after incubation and subsequent staining of the gel the band corresponding to the r-silicatein protein was unstained (Fig. 5B, lane a), indicating that the gelatin was digested at that position. This result indicates that r-silicatein also has proteolytic activity.

Inhibition studies were performed to support this conclusion. As outlined before (Krasko et al., 1997), following the protocols described (Beynon and Bond, 1989), the zymograms were incubated in Tris buffer, supplemented with the cysteine protease inhibitors *p*-chloromercuriphenylsulfonic acid (1 mM), iodoacetic acid (1 mM), *N*-ethylmaleimide (1 mM), E-64 (10 µM;

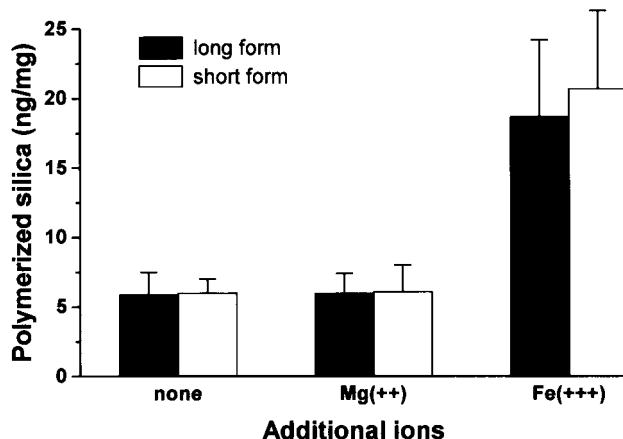


Fig. 4. Dependence of the activity of silicatein on $\text{Fe}^{(++)}$. The assay was performed with the two r-silicatein preparations (long as well as short form) using 4.5 mM tetraethoxysilane (TEOS) as substrate and, where indicated, MgSO_4 or $\text{Fe}^{(++)}$ citrate. After incubation the amount of polymerized silica was quantified after treatment with NaOH; the released silicic acid was determined ($n = 5$; $\pm \text{SD}$). The values are given in ng of silicon (polymerized), and if enzyme was added, correlated with 1 mg of protein; means of five separate experiments are presented.

Fig. 5C, lane c) or cystatin (1 mg/ml; lane b). All inhibitors reduced the proteolytic activity of silicatein in the zymogram assay. From these data we concluded that silicatein has the characteristic activity of cysteine peptidase(s).

BIOLOGICAL ROLE OF SILICON Localization in Sponges

Silica is stored in sponges in the form of spicules. Two intracellular compartments which contain silica have been identified: the extranuclear and the intranuclear one.

Spicules. The spicules are formed by scleroblasts (reviewed in Simpson, 1984). The formation starts around an axial filament (Fig. 6A). Spicules can grow at up to 5 $\mu\text{m}/\text{h}$ (Weissenfels, 1989). The first microscopical study was performed by Lieberkühn in 1856 (Fig. 6B).

Nucleus. The fact that siliceous spicules also occur in the nuclei of cells of the freshwater sponge *Spongilla lacustris* (Fig. 6D) was surprising (Imsiecke et al., 1995). Digestion studies showed that the crystals, observed by Lieberkühn (1856) (Fig. 6C), consist of silica; they are not digested by proteinase K but by hydrofluoric acid (Imsiecke et al., 1995). The physiological role of the intranuclear silicon rods is unknown. Since they show a high metabolic rate they are assumed to be involved in the scavenging of other ions or in the maintenance of osmotic pressure within the cell (Imsiecke et al., 1995).

Effect on Size of Primmorphs

Primmorphs. Primmorph cultures have been established from the demosponges *S. domuncula* (Custodio et al., 1998; Müller et al., 1999), *Dysidea avara* (Müller et al., 2000), and occasionally *G. cydonium*. In brief, tissue samples were dissociated to single cells in Ca^{2+} -

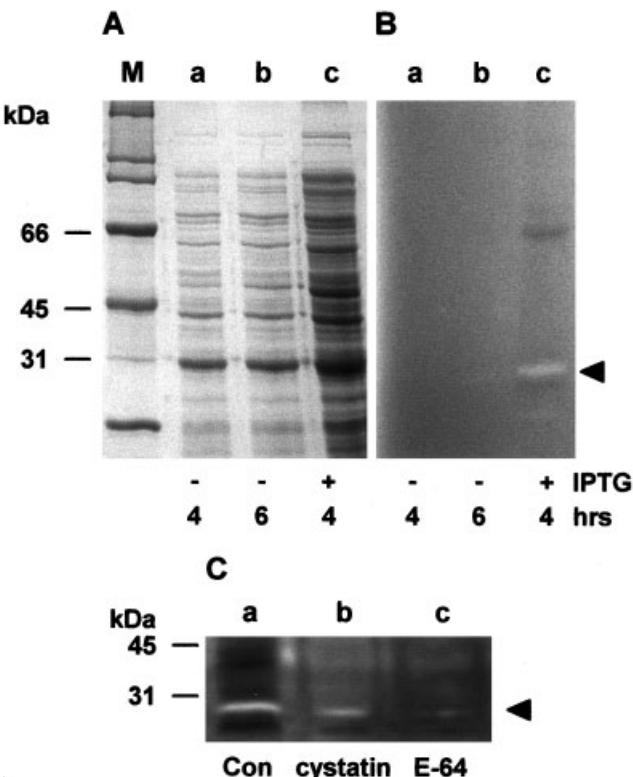


Fig. 5. Demonstration of the proteolytic activity of silicatein. $\text{NaDdSO}_4/\text{PAGE}$ (10% gel) was performed in the presence of NaDdSO_4 . The gels were prepared in the absence (A) or together with 0.1% gelatin (Sigma, St. Louis, MO; G-2500) (B). Bacterial extracts, after expression of the short form of silicatein (see Fig. 3), were applied to the gels at a concentration of 20 μg per lane. The samples were obtained either from bacteria which had been induced with IPTG for 4 hours (+IPTG; lane a) or were cultivated in the absence of this inducer for 4 or 6 hours (-IPTG; lanes b, c). A: The gels which did not contain gelatin were immediately stained with Coomassie brilliant blue; in parallel, protein markers were separated. B: Alternatively, the bacterial samples were applied in the same order onto the gelatin-containing gels and separated. After the run the gels were washed in Tween-20 solution and twice in a 100 mM Tris/HCl buffer (pH 8.0; 0.01 M CaCl_2). Finally, the gels were incubated for 18 hours in this Tris buffer at 20°C and then stained with Coomassie brilliant blue. The arrow indicates the location where the gelatin had been digested and remained unstained by Coomassie blue. C: Inhibition studies. The extracts from bacteria which had been induced with IPTG were applied at equal concentrations to the three slots and electrophoresed; lanes a–c. After size separation the zymograms were cut into slices and incubated separately in the above-mentioned 100 mM Tris/HCl buffer in the absence of any further component (lane a), or in this buffer supplemented with 1 mg/ml of cystatin (lane b) or 10 μM of E-64 (lane c). Then the zymograms were stained in Coomassie brilliant blue.

and Mg^{2+} -free artificial seawater containing EDTA. Subsequently, the cells were transferred to natural seawater and kept at 16°C. When aggregates formed, they were incubated in seawater containing 0.2% of RPMI1640-medium (Le Pennec et al., 2003), where they form primmorphs.

The primmorphs are defined by the presence of proliferating cells and a characteristic histology (Custodio et al., 1998; Müller et al., 1999). During the phase of primmorph formation the aggregates contract to round-shaped bodies, 1–5 mm in diameter, leaving be-

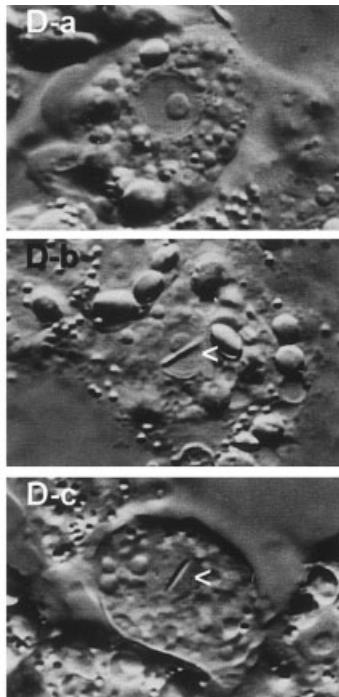
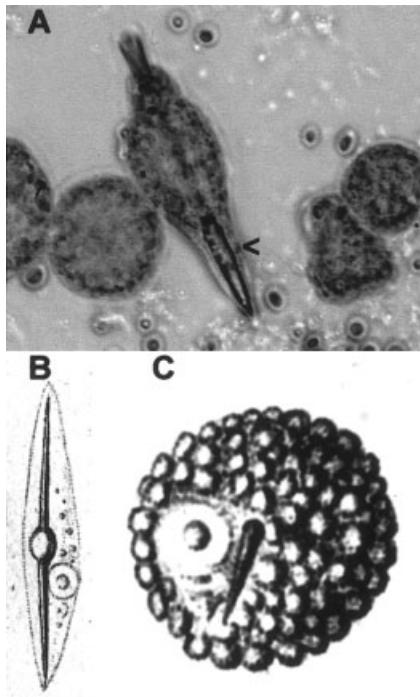


Fig. 6. Formation of spicules in the freshwater sponges *Ephydatia muelleri* and *Spongilla lacustris*. **A:** Formation of the spicules in scleroblasts from *E. muelleri* (magnification $\times 1,000$; Imsiecke and Müller, unpubl.). The spicules (<) start to grow around an axial filament. **B:** Drawing showing the initiation of the spicule formation within a scleroblasts (Lieberkühn, 1856). **C:** The intracellular presence of a spicule in the neighborhood of the nucleus of the sponge cell (Lieberkühn, 1856). **D:** Formation of a siliceous spicule in the nucleus of *S. lacustris* ($\times 1,200$). The growing crystal (<) is shown (modified after Imsiecke et al., 1995).

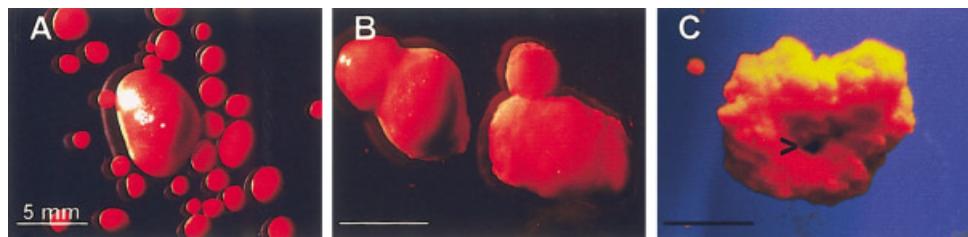


Fig. 7. Formation of primmorphs from dissociated cells of *S. duncula*. **A:** When the cells are transferred to seawater, supplemented with 0.2% of RPMI1640, primmorphs of a size between 1–7 mm are formed (incubation period 5 days). The large-size primmorphs (one specimen is shown) are very rare. **B:** Incubation for 5 days of the primmorphs in this medium, supplemented with 60 μM silicate and 30 μM Fe(++) (as ferric citrate); size of the primmorphs ranges

from 6–12 mm. **C:** Incubation of the primmorphs in seawater/RPMI1640/silicate/30 μM Fe(++) in the current chamber at a water velocity of 20 cm/sec. The primmorphs under these conditions often show an oscule-like opening (arrowhead). Scale bar = 5 mm. [Color figure can be viewed in the online issue, which is available at www.interscience.wiley.com]

hind detritus and dead cells. In the initial phase the primmorphs remain round-shaped but after 3–4 weeks many of them attach to the bottom of the culture dish. Cross sections through the primmorphs revealed that the inner cells are surrounded by an almost complete single-cell layer of epithelial-like cells. The cells forming the epithelium of the primmorphs are pinacocytes, as judged from their flattened, fusiform extensions and their prominent nucleus. In the absence of any additional factor, the diameter of the primmorphs ranged from 1–7 mm (Fig. 7A); the usual diameter of the round-shaped primmorphs is ~4 mm.

The growth conditions were optimized by supplementing the natural seawater, including 0.2% of RPMI1640-medium, with silicate and Fe(++). Furthermore the primmorphs were kept in a current chamber (Le Pennec et al., 2003). If the culture medium was enriched with 60 μM silicate and 30 μM Fe(++), the primmorphs grew; 6–12 mm-long 3D-

aggregates were usually obtained (Fig. 7B). When the incubation was performed under higher water current, the primmorphs frequently formed oscule-like openings (Fig. 7C).

Morphogenetic Activity. Two inorganic factors have morphogenetic activity on primmorphs: silicate and ferric iron.

Effect of Silicate. In a preliminary series of experiments the effect of silicate on spicule formation in vitro was determined semiquantitatively (Krasko et al., 2000, 2002). When the primmorphs were incubated in the absence of exogenous silicate for 5 days no spicules were seen. In contrast, when the primmorphs were incubated together with 50–250 μM of silicate, bundles of spicules were found. The content of polymerized silicon also increased in primmorphs in the presence of silicate. In the absence of additional silicate in the medium the primmorphs contained little polymerized silicon (0.1 mg/0.4 g aliquots⁻¹); however, when they

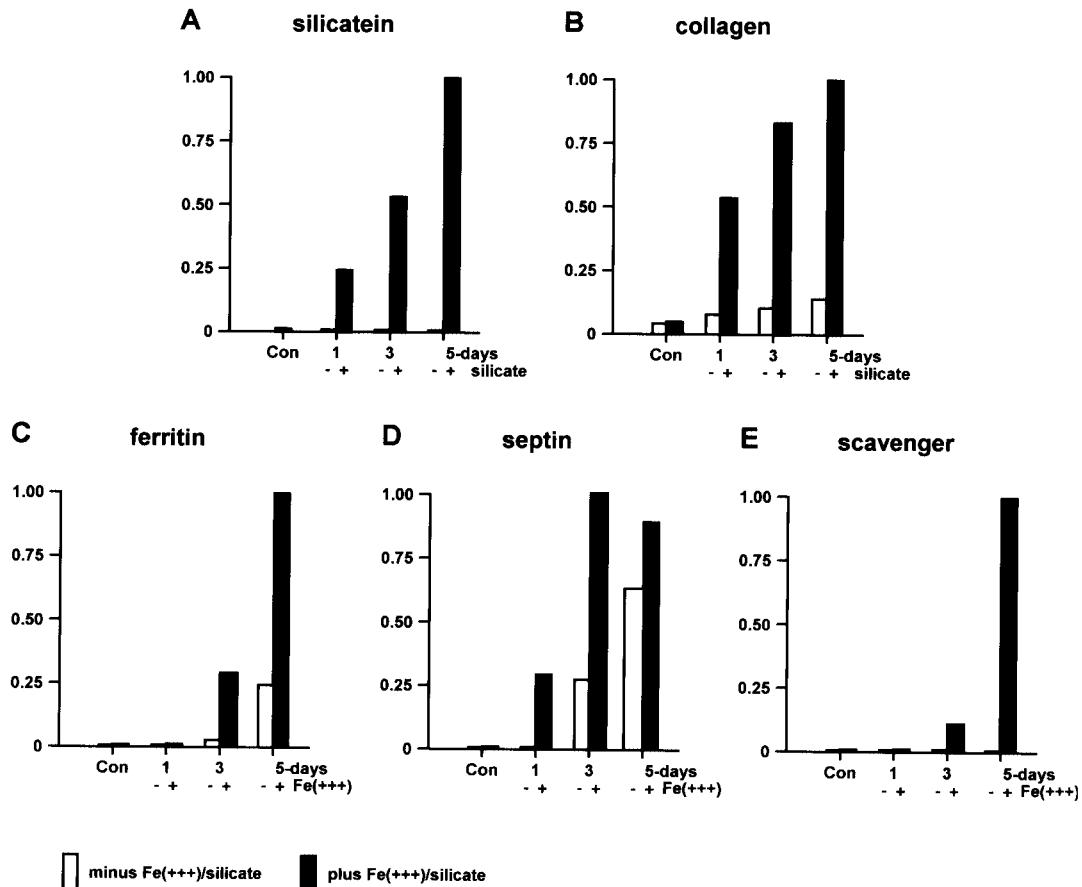


Fig. 8. Effect of silicate and ferric iron on the steady-state expression level of selected genes in primmorphs. Single cells were transferred after dissociation into Ca^{2+} and Mg^{2+} -free artificial seawater containing EDTA into seawater, enriched with 0.2% of RPMI1640. During primmorph formation the cultures remained either without any additional component for 0 (control; "Con") to 5 days or they were incubated together with silicate 60 μM silicate (A,B). In a further series, the seawater/RPMI1640/silicate medium was additionally supplemented with 30 μM Fe(++) (C-E). Primmorphs were collected, RNA was extracted, and the level of transcripts was estimated by Northern blot analysis, as described previously (Krasko et al., 2000,

were cultured in seawater medium enriched with 60 μM silicate the content increased to 4.4 mg.

Several genes are expressed before or during spicule formation (Krasko et al., 2000, 2002). The effect of silicate on the steady-state expression level of silicatein and collagen in primmorphs has been described (Krasko et al., 2000). The genes for silicatein and collagen were cloned and used for Northern blot studies.

The concentration of silicate in the natural seawater used for the experiments was $\approx 1 \mu\text{M}$. In this medium the expression of silicatein in primmorphs is hardly detectable (Fig. 8A). However, when the concentration of silicate was raised to 60 μM , the expression of silicatein strongly increased after 1 day, to reach a maximum after 5 days (Fig. 8A). Furthermore, the expression of the gene encoding collagen (Schröder et al., 2000) also responded to silicate (Fig. 8B). In natural seawater the expression of collagen was relatively low and amounted to $\approx 15\%$ of that seen in the presence of 60 μM exogenous silicate.

2002). The following cDNAs were used as probes: (A) silicatein, (B) collagen, (C) ferritin, (D) septin, or (E) scavenger. After blotting, the intensities of the hybridized bands were semiquantitatively determined using the GS-525 Molecular Imager (Bio-Rad, Hercules, CA). The relative degree of expression was correlated with that seen for the maximal expression after 5 days in the presence of silicate, which was set to 1.0 (A,B). In the studies to estimate the effect of Fe(++) (C-E), the values are correlated with the expression at day 5 in primmorphs cultured in the presence of silicate and Fe(++); modified according to Krasko et al. (2002).

Effect of Ferric Iron. As outlined above, addition of Fe(++) results in the formation of larger primmorphs (Fig. 7).

In addition, ferric iron affects the expression of the following genes: ferritin, the iron storage protein (reviewed in Chasteen and Harrison, 1999), a putative septin polypeptide, and a scavenger receptor. Septin molecules are involved in the completion of cytokinesis (reviewed in Xie et al., 1999). Since the spicules in some Demospongiae are formed extracellularly (Uriz et al., 2000) it has been assumed that cell surface receptors are involved in the growth guidance/control of the siliceous spicules (unpublished). However, the expression of the sponge integrin receptor did not depend on extracellular iron, while a considerable upregulation of a scavenger receptor was observed after addition of iron to primmorph cultures. This scavenger receptor was also cloned (Krasko et al., 2002).

The effect of iron on the expression level of these genes, ferritin, septin, and scavenger receptor, in prim-

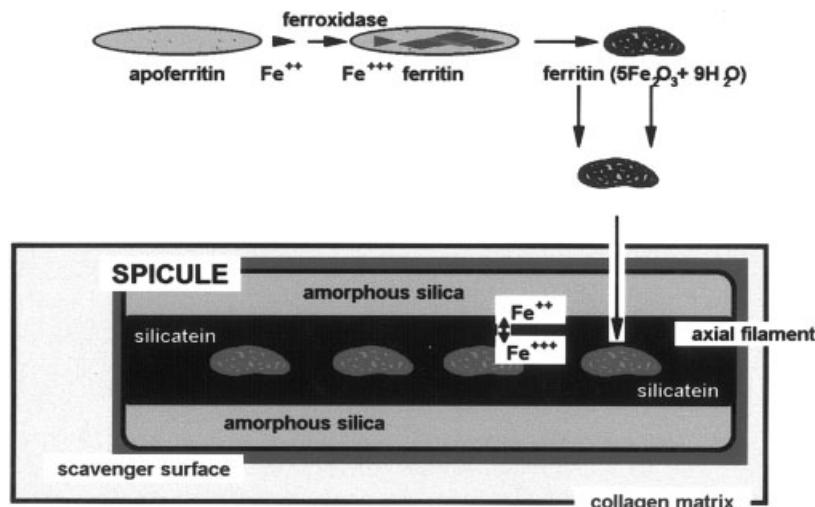


Fig. 9. Proposed biochemical steps involved in the spicule formation in Demospongiae. It can be suggested that Fe^{++} is taken up by sponge cells and undergoes oxidation to Fe^{+++} via the ferroxidase of ferritin. Ferritin/ Fe^{+++} is translocated to the axial filament, where it is coated by silicatein. This enzyme, which is mediating the formation of amorphous silica, is very likely activated by iron. This skeletal element, amorphous silica, is surrounded by the scavenger receptor and finally embedded into the collagen matrix.

morphs was also determined by Northern blot experiments. Primmorphs were transferred to seawater and supplemented with 60 μM silicate and 30 μM Fe^{+++} . Without Fe^{+++} the expression of the 0.9 kb *ferritin* increased slightly after 5 days (Fig. 8C). In contrast, if RNA was isolated from primmorphs which had been cultivated in the presence of Fe^{+++} , *ferritin* expression increased after 3 days, and continued to rise for 5 days. In parallel, samples were hybridized with the *septin* probe. Again, the expression of the *septin* gene strongly increased in cultures treated with Fe^{+++} , but not in its absence (Fig. 8D). In addition, increased *scavenger receptor* expression was seen after exposure to iron for 3 and 5 days, but not in the iron-free controls (Fig. 8E).

These data indicate that, besides silicate, iron is also essential for proliferation and morphogenesis of sponge primmorph cells. This view was also substantiated by *in situ* hybridization studies (Le Pennec et al., 2003). Evidence showed that in the initial phase of reorganization iron enhances proliferation followed by a morphogenetic effect via its accelerative and supportive influence on spicule synthesis.

CONCLUSION

It is well established that silicon and iron are major elements in sponge spicules (Vinogradov, 1953). Following the studies performed with *T. aurantium* (Cha et al., 1999) and *S. domuncula* (Krasko et al., 2000), it appears to be well established that silicatein is the major enzyme involved in spicule formation.

Applying the primmorph system from *S. domuncula*, the data (Krasko et al., 2002) suggested that besides silicon, iron ions play an important role in the initiation of spicule formation. Iron upregulates ferritin in sponge cells, which may bind the intracellular Fe^{++} . Subsequently, Fe^{++} is oxidized to Fe^{+++} under consumption of oxygen (Ha et al., 1999); this process is mediated by the ferroxidase site of ferritin. During this process apoferitin is converted to ferritin. Finally, iron may be deposited in ferritin as $5\text{Fe}_2\text{O}_3 \cdot 9\text{H}_2\text{O}$ and then as the stable mineral $\alpha\text{-Fe}_2\text{O}_3$ (see Chasteen and Harrison, 1999). The deposited mineralized iron, the ferric

oxide nanoparticles, remain surrounded by the ferritin protein shell. In the iron-free state the ferritin molecules (apoferitin) comprise hollow spheres with an outer diameter of 12 nm and an inner diameter of 8 nm (Lawson et al., 1991). Therefore, ferritin can be considered as a shell that causes both the oxidation of iron ions and controls the growth of the iron mineral.

Recent studies applying the technique of *in situ* hybridization with *S. domuncula* revealed that ferritin is abundant in cells surrounding the canals within the sponge body (Le Pennec et al., 2003). The function of ferritin in those cells remains to be established. Based on analytical (Vinogradov, 1953) and functional data (Krasko et al., 2002) the deposition of iron in ferritin spheres may also be involved in the initiation of spicule formation (Fig. 9). Since the diameter of the axial filament, which also comprises the silicatein, is $\approx 1 \mu\text{m}$ (Uriz et al., 2000) there is ample space for the ferritin shell. It may be assumed that these spheres may be covered by silicatein, which results in the initiation of spiculogenesis. Silicatein is finally activated by iron [as Fe^{+++}]. This assumption is supported by the data, which indicate that Fe^{+++} stimulates the activity of silicatein. In addition, iron within the ferritin shell is mobilized after reduction (Chasteen, 1998). Another gene, coding for a scavenger receptor, is apparently involved in the spicule formation. The expression studies suggest that after or during the synthesis of amorphous silica, the growing spicules are surrounded by the scavenger receptor, which might be considered a docking molecule for the collagen matrix into which the spicules are embedded (Fig. 9).

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